



STATE OF NEW YORK  
DEPARTMENT OF HEALTH  
*Virology Proficiency Testing Program*

## General Category

May 5, 2009

Summary of scores, responses, and  
statistics for General Category Samples:  
1111, 1112, 1113, 1114, 1115

**Disclaimer**

**The use of brand and/or trade names in this summary does not  
constitute an endorsement of the products on the part of the  
Wadsworth Center or the  
New York State Department of Health**

**The New York State Proficiency Testing Program May 2009 General Category Evaluation Reports are available at:**  
<https://commerce.health.state.ny.us/doh3/applinks/epters/> and can be printed for your records.

This summary is based on scores and responses in the Electronic Proficiency Testing Reporting System.

### **Sample Description:**

The May 2009 General panel contained four simulated patient specimens that were isolates from patient specimens and one no virus sample. All NYSDOH Proficiency Test samples were isolates of viruses grown in the Virus and Surveillance Laboratory at the Wadsworth Center.

**NYS PT Sample 1111, Influenza virus, type B**, was a simulated nasal wash from a three year old male. The virus was originally isolated in 2007 from a NPS from a three year old male with fever and an upper respiratory infection. The virus was determined to be B/OHIO/01/2005-LIKE by the CDC.

**NYS PT Sample 1112, Adenovirus 21**, was a simulated NPS from a 28 year old female. The virus was originally isolated in 2006.

**NYS Sample 1113, Herpes simplex virus, type 1**, was a simulated vesicle swab from a 21 year old female with fever and a skin lesion. The virus was originally isolated in 2006.

**NYS Sample 1114** was a **No virus** specimen.

**NYS Sample 1115, Coxsackievirus, type B3**, was a simulated CSF. The virus was originally isolated from an isolate in RhMk of a CSF from a 27 year old female with a frontal headache in 2006.

### **Sample Scoring and Validation**

The scores and analysis from the May 2009 General Panel are shown below. Federally mandated validation criteria require a sample to be correctly identified by at least 80% of participating laboratories. All five samples are valid and their identities were also confirmed by reference laboratories. CLIA and CLEP establish a passing grade for participating laboratories at 80% or greater.

**Please be aware that scoring is based on the number of samples your facility tested. No credit will be give for samples not tested.** For example, if a facility tested four of the five PT samples, the total score would be based on four responses, each worth, 25%. Therefore, if one response was incorrect, the total score would be 75%, a failing grade.

## May 2009 General Event

### Scoring Analysis: 43 Participating Laboratories

Sample #	1111	1112	1113	1114	1115
Sample Identification	Influenza virus, type B	Adenovirus, type 21	Herpes simplex virus, type 1	No virus	Coxsackie virus, type B3
Titer (TCID <sub>50</sub> ) Log 10/ml	6.5	3.7	6.5	0	8.5
Laboratories Scoring 100%	43	42	43	43	43

### General Grade Distribution

Total Score For Panel	100%	80%	60%	40%	20%	0%
Participating Laboratories	42	1	0	0	0	0

### Culture Methods Reported by Laboratories

	# of Responses Using Each Method/Sample				
	Sample 1111	Sample 1112	Sample 1113	Sample 1114	Sample 1115
Conventional	16	16	17	16	26
Centrifugation Enhanced	7	7	6	4	3
Conventional & Centrifugation Enhanced	20	20	18	23	14

**Cell lines utilized by participating laboratories for culture of each sample\***

<b>Cell Lines</b>	<b>1111 Inoculated</b>	<b>1111 Detected In</b>	<b>1112 Inoculated</b>	<b>1112 Detected In</b>	<b>1113 Inoculated</b>	<b>1113 Detected In</b>	<b>1115 Inoculated</b>	<b>1115 Detected In</b>
A549	12	1	19	19	20	20	19	9
AGMK	1		1	1	1	1	1	1
BGMK							1	1
CV-1			1		1	1	1	
E-Mix	3	1	3	1	3	1	9	9
ELVIS					4	3		
H & V Mix					5	4	1	
HEL	1		1	1			1	1
Hep-2	3		5	4	2	1	4	3
HFF	2		2	1	1	1	3	
ML					1			
MKCy	1	1	1				1	1
MRC-5	15	1	19	12	30	26	23	6
MRHF	1	1						
pRK					4	4		
R-Mix	12	12	11	11	1	1	1	1
R-Mix Too	4	4	5	4			1	
RhMK	29	28	21	12	10	3	37	34
Super E-Mix							1	1
Vero					2	2	1	1
WI-38	1		2	1	0			

\*To generate this data, the first two selections listed on the submitted information have been captured from each laboratory. The additional selections by laboratories that inoculated more than two cell lines have not been included.

“Cell Line Inoculated” or “Detected In” data is not included in the above chart for Sample 1114, No virus.

# Sample 1111

## Influenza B

### Confirmation Reagents

Reagent Name	# of Responses
Bartels	2
Bartels VRK	1
Binax NOW Influenza A & B	1
D3 Ultra DFA Respiratory Virus & Screen ID kit	18
DAKO Imagen Influenza Kit	2
DHI	3
DHI Influenza B DFA	2
Light Diagnostic	1
Light Diagnostic Respiratory Panel	1
Millipore	1
Millipore Respiratory DFA Screen kit	2
PathoDX Influenza A	1
Prodesse ProFlu+	1
Quidel QuickVue	1
Quiagen	1
Respiratory virus ID DFA kit	1
Simulfluor Flu A/B	2

### Confirmation Methods

Method	#of Responses
EIA	3
HAD	2
Immunofluoresence	36
PCR	2

**Sample 1112**  
**Adenovirus, type 21**

**Confirmation  
 Reagents**

Reagent Name	# of Responses
Adenovirus DFA kit	1
Bartels	2
Bartels VRK	1
Bartels MAB	1
Chemicon Light Diagnostics	5
DAKO Imagen	1
DHI	3
DHI Adeno MAB	1
DHI Adenovirus DFA	2
D3 Ultra DFA Respiratory Virus & Screen ID kit	16
Light Diagnostic	1
Light Diagnostic Adeno MAB	1
Light Diagnostic Respiratory Panel	1
Millipore	1
Millipore Respiratory DFA Screen kit	2
PathoDX	1
Respiratory Viral Panel FA kit	1
Respiratory virus ID DFA kit	1

**Confirmation Methods**

Method	# of Responses
EIA	1
Immunofluoresence	41

**Sample 1113**  
**Herpes simplex virus, type 1**

**Confirmation Reagents**

Reagent Name	# of Responses
Bartels	3
Bartels HSV Bivalent DFA kit	2
Bartels HSV Typing	1
Chemicon Simulfluor HSV/VZV DFA	1
D3 DFA HSV ID/Typing kit	2
DHI D3 DFA HSV ID/typing kit	1
DHI ELVIS Typing kit	3
DHI HSV ID kit	1
HSV 1/HSV 2 Culture ID Typing	1
Light Diagnostic	2
MicroTrak	1
MicroTrak HSV ID kit	4
Millipore HSV Culture ID	1
Millipore Light Diagnostics HSV Reagent	2
PathoDX	5
PathoDX Herpes Typing	1
Simulfluor HSV 1/2	2
SYVA Trinity	1
Trinity Biotech HSV ID/Typing	3
Trinity Biotech MicroTrak	3
Trinity Herpes Bivalent kit	1

**Confirmation Methods**

Method	# of Responses
ELVIS	2
Immunofluorescence	35

**Sample 1115**  
**Coxsackievirus, type B3**

**Confirmation  
 Reagents**

Reagent Name	# of Responses
Chemicom	3
Chemicon Coxsackie B DFA	1
Chemicon Light Diagnostics	1
DHI	2
DHI Enterovirus ID kit	2
DHI D3 IFA Enterovirus kit	2
Enterovirus PCR	1
Homebrew	1
Light Diagnostics	6
Light Diagnostics Cox B MABS	1
Light Diagnostics Pan-Entero	4
Millipore	3
Millipore Coxsackie Antibody	1
Millipore Enterovirus Screen	1
Pan-Entero Blend	1
Pan-enterovirus Screen	1
Pan-Entero FA, Cox B3 FA	1

**Confirmation Methods**

Method	# of Responses
Immunofluoresence	37
PCR	2

Confirmation Reagent data is not included in the above charts for Sample 1114, No virus.

## Notes:

- The September 2009 panel will contain one **EDUCATIONAL SAMPLE**. It is not mandatory that you test this sample-it is an **optional** sample. Your response for this sample can be entered in EPTRS. The Evaluation Report will list your laboratory's result, a feedback response (correct vs. incorrect) as well as the consensus response. **This educational sample will not be included in your final grade**. Please notify staff about this additional sample.
- The NYS Virology Proficiency Testing laboratory will be re-located during the month of October 2009. Our mailing address will remain the same. However, we will have new phone and fax numbers. You will receive further information about new contact numbers as soon as it is available. We would also like to inform you that the Virology PT shipping schedule for 2010 may have slight differences than in the past.
- If you are using a confirmation method that detects and types a virus, it is expected that you will report the virus type.
- Lot numbers and expiration dates for Confirmation Reagents/Kits are no longer a required field.
- Generic worksheets are provided for your optional use. Please do not return these worksheets to us. Generic worksheets can also be printed via the HPN (<https://commerce.health.state.ny.us/doh3/applinks/eptrs/>).
- Participants **MUST** enter responses in ALL fields when reporting electronically; scores may be adversely affected if there are blank fields.
- In the "Virus Isolated" drop down menu, please select "No virus" if your intended answer is "No virus isolated." The scoring algorithm does not recognize "No virus isolated" and your score may be affected. In the case where "No virus" is selected, please select "Not Applicable: no virus isolated" in the Cell Line Detected In field. For the field, Confirmation Method, select No Confirmation Testing Performed; and in the field, Confirmation Reagents, type NA.

## EPTRS Notes:

- Participation in EPTRS is mandatory. Laboratories must submit test results electronically by logging in to: <https://commerce.health.state.ny.us/doh3/applinks/eptrs/> entering their results and clicking the "Submit/Attest" button on the EPTRS Summary Page.
- Please be sure to "Submit" test results. Keeping results as "Saved" is considered non-participation for the event and will automatically result in a failing grade in the electronic system.