



ANDREW M. CUOMO
Governor

**Department
of Health**

HOWARD A. ZUCKER, M.D., J.D.
Commissioner

SALLY DRESLIN, M.S., R.N.
Executive Deputy Commissioner

**New York State Department of Health - Wadsworth Center
Laboratory of Organic and Analytical Chemistry–
NYS ELAP Laboratory ID 10763**

**Division of Environmental Health Sciences
Albany, New York**

**Medical Marijuana sample preparation protocols for potency analysis
NYS DOH MML-301**



ANDREW M. CUOMO
Governor

Department of Health

HOWARD A. ZUCKER, M.D., J.D.
Commissioner

SALLY DRESLIN, M.S., R.N.
Executive Deputy Commissioner

Table of Contents

1.0.	Scope and Application	3
2.0.	Summary of the Method.....	3
3.0.	Definitions.....	3
4.0.	Health and Safety Warnings.....	5
5.0.	Interferences.....	5
6.0.	Equipment and Supplies	6
7.0.	Reagents, Standards and Matrix (Consumables).....	7
8.0.	Preparation of Reagents, Solutions, Standards, Matrices and Samples.	8
9.0.	Quality Control/Assurance	9
10.0.	Data Acquisition, Reduction, Analysis and Calculations	10
12.0.	Waste Management/Pollution Prevention	11
13.0.	References.....	11



ANDREW M. CUOMO
Governor

Department of Health

HOWARD A. ZUCKER, M.D., J.D.
Commissioner

SALLY DRESLIN, M.S., R.N.
Executive Deputy Commissioner

1.0. Scope and Application

- 1.1. This method (NYS ELAP Method ID 9981) only addresses the extraction of medical marijuana (MM) samples for cannabinoid analysis by High Performance Liquid Chromatography (HPLC) with Photodiode Array (PDA) Detection. It contains information specifically relevant to the extraction and preparation of MM products in **Section 8.0**. Refer to the analytical procedure (**NYS DOH MML-300**) for information on analyte list, calibration, analysis, quality control and data reporting.
- 1.2. This method is restricted to use by or under the supervision of analysts experienced in the preparation of MM products. Each analyst must demonstrate the ability to generate acceptable results with this method using the procedures described in **Section 9.0**.

2.0. Summary of the Method

- 2.1. A portion of MM product, typically from 10 to 1200 mg, is weighed into a 50-mL centrifuge tube. The amount weighed depends upon the specific product produced by a Registered Organization (RO) and the declared concentrations of cannabinoids in the MM product. A surrogate (SUR) and 20.0-mL of methanol (MeOH) are added, the solution is mixed well and is either diluted further or used directly for analysis. If necessary, this extract is diluted an additional 2- to 20-fold based on the concentrations of cannabinoids in the MM sample as declared by the RO. The internal standard working diluent (IWD) is then added to the extract or dilution thereof, and the potency measurement is made using HPLC-PDA (**NYS DOH MML-300**).

3.0. Definitions

- 3.1. Stock Standard – A concentrated solution of method analyte(s) prepared in the laboratory from referenced and certified analyte standards, where available, or a concentrated solution of method analyte(s) purchased directly from a referenced and certified source, where available.
- 3.2. Internal Standard (IS) – A pure compound that should not be found in any sample. The IS is a compound added to both samples and standards at a known concentration to provide a basis for peak area ratios used in quantitation. The IS is also used to monitor instrument performance for each analysis and to correct for solvent evaporation during the analysis.
- 3.3. Internal Standard Stock Diluent (ISD) – A concentrated solution of IS that is prepared in solvent. This stock diluent is used to prepare the internal standard working diluent (IWD).
- 3.4. Internal Standard Working Diluent (IWD) – A solution of IS that is prepared from the ISD and added to all samples at the same concentration. This working diluent is used to dilute the samples and monitor the integrity of the sample injections.
- 3.5. Surrogate Standard (SUR) – A pure analyte, which should not be found in any sample, but is similar in nature to the compounds of interest. This compound can be added to a sample in a known amount before processing to monitor method performance for each sample.
- 3.6. Surrogate Stock Diluent (SSD) – A concentrated solution of SUR that is prepared in acetonitrile (MeCN). This stock diluent is used to prepare the surrogate working diluent (SWD).
- 3.7. Surrogate Working Diluent (SWD) – A solution of SUR that is prepared from the SSD and is added to all samples. This working diluent is used to monitor method performance.
- 3.8. System Blank (SBLK) – A portion of appropriate pure solvent that is analyzed to verify that the



instrument is free from background contamination.

- 3.9. Method Blank (MB) – An aliquot of appropriate pure matrix that is treated exactly as a sample including exposure to all glassware, equipment, solvents, reagents and surrogates that are used with other samples. The method blank is used to determine whether method analytes or other interferences are present in the laboratory environment, reagents or apparatus.
- 3.10. Continuing Calibration Verification Standard (CCV) – One of the primary calibration standards used to verify the acceptability of an existing calibration.
- 3.11. Cross Check Reference Standard (CCR) – A solution of method standards prepared from a stock standard solution that is obtained from a source that is independent of that used to prepare the calibration standards (i.e. independent vendor, independent lot, or independent preparation). The CCR is used to verify that the original calibration source is acceptable
- 3.12. Laboratory Control Sample (LCS) – A portion of appropriate clean matrix that is spiked with known quantities of target analytes and processed as a sample. The LCS measures the accuracy of the methodology. Acronyms include: Method Blank Spike (MBS) and Laboratory Fortified Blank (LFB). A CCV or CCR may also serve as an LCS for this procedure. A separate LCS does not have to be included in an analytical batch when either of the designated CCV or CCR samples meets the LCS criteria.
- 3.13. Matrix Spike Sample (MS) – A portion of sample that is spiked with known quantities of target analytes and processed as if it were a sample. The sample from which the portion to be spiked was taken must be analyzed separately to determine any background analyte concentrations. The MS is corrected for background concentrations and used to determine whether the sample matrix contributes bias to the sample results. The MS is used to evaluate the accuracy of the method in the same way that the MBS is used.
- 3.14. Matrix Spike Duplicate Sample (MSD) – A second portion of the sample that was used to prepare the MS that is spiked and processed in an identical manner to that used for the MS. The MS and MSD are used together to measure the precision of the method.
- 3.15. Limit of Detection (LOD) – The statistically calculated minimum concentration of an analyte that can be measured with 99 % confidence that the value is greater than zero. Acronym: Method Detection Limit (MDL).
- 3.16. Limit of Quantitation (LOQ) – The minimum concentration that can be quantitatively reported for a target analyte. This limit can be no lower than the lowest calibration standard.
- 3.17. Sample Batch – A group of Cannabinoid extracts that are processed together as a unit using the same procedure and materials. A typical batch consists of 20 samples. A batch includes applicable quality controls, such as method blanks, method blank spikes, matrix spikes, duplicates, or quality control samples. (Batch ID#).
- 3.18. Analysis batch – A set of samples that are analyzed on the same instrument during a 24-hour period



ANDREW M. CUOMO
Governor

Department of Health

HOWARD A. ZUCKER, M.D., J.D.
Commissioner

SALLY DRESLIN, M.S., R.N.
Executive Deputy Commissioner

4.0. Health and Safety Warnings

- 4.1. The toxicity and carcinogenicity of each chemical used in this method have not been thoroughly investigated. Therefore, each chemical compound must be treated as a potential health hazard and exposure must be limited to the lowest possible level.
- 4.2. Always follow guidelines listed in safety data sheets (SDS) for proper storage, handling and disposal of solvents, reagents and standards. These guidelines must be made available to all personnel involved in the chemical analysis.
- 4.3. Lab coats, safety glasses and gloves must be worn when performing standard or sample preparations, working with instrumentation, disposing of waste and cleaning glassware.
- 4.4. The fume hood must be used when using or preparing standards, reagents, or samples that require proper ventilation.
- 4.5. The IS norgestrel is a suspected carcinogen and is known to be hazardous during pregnancy.

5.0. Interferences

- 5.1. Method interferences may be caused by contaminants in solvents, reagents, glassware and other sample processing apparatus that lead to discrete artifacts observed as chromatographic peaks or elevated baselines in the chromatograms. All reagents and apparatus must be routinely demonstrated to be free from interferences under the conditions of the analysis by running extracted blanks as described in **NYS DOH MML-300**.
- 5.2. All glassware must be washed and, if applicable, verified to be free from background contamination.
 - 5.2.1. All new glassware and processing apparatus must be thoroughly cleaned. Before using new glassware or equipment the first time, wash with hot water and detergent, rinse with tap water and reagent water and final rinsing with methanol.
 - 5.2.2. All routine glassware and processing apparatus must be thoroughly cleaned. After each use, rinse all glassware and processing apparatus three times with the last solvent used and dry in a clean area to prevent cross-contamination. If glassware contamination is suspected wash as per **Section 5.2.1**.
 - 5.2.3. The use of high-purity reagents and solvents helps to minimize interference problems.
 - 5.2.4. After cleaning, glassware is stored in a clean storage area away from standards and syringes to prevent cross-contamination.
- 5.3. When interferences or contamination are evident in samples, the re-preparation of the original sample is recommended after the source of contamination has been identified.
- 5.4. Interfering contamination known as “carry over” may occur when a sample containing low concentrations of analytes is analyzed immediately following a sample containing relatively high concentrations of analytes. Rinsing of the sample syringe and associated equipment between samples with solvent/mobile phase can minimize this sample cross contamination. After analysis of



a sample containing high concentrations of analytes one or more injections of solvent/mobile phase should be made to ensure that accurate values are obtained for the next sample.

- 5.5.** Matrix interferences may occur because of contaminants present in the sample. If a matrix interference is believed to be present, a matrix-spike experiment must be performed. The MS must be analyzed together with the un-spiked sample to verify the results. If these analyses verify the original results, report only the results from the original un-spiked sample. This may not always be possible if a limited amount of sample is received for analysis. If additional sample is not available for reanalysis, the original results must be qualified on the final report.
- 5.6.** Samples and standards must be prepared in the same final solvent to allow for chromatographic comparability of samples to standards.

6.0. Equipment and Supplies

6.1. Sampling Equipment

- 6.1.1.** Pre-cleaned 50-mL plastic bottle fitted with Teflon-lined screw cap.

6.2. Equipment

- 6.2.1.** Analytical balance, Mettler-Toledo Model # 205DU or equivalent (DEHS-008 Weight/Balance SOP)
- 6.2.2.** Sonicator, Branson, Model # 2510R-DTH or equivalent.
- 6.2.3.** Vortex, Maxi Mix 11 Model #37615 or equivalent.
- 6.2.4.** Centrifuge, Model # 5415D or equivalent.
- 6.2.5.** Shaker, Labline, Model# 3540 or equivalent.

6.3. Support Equipment

- 6.3.1.** Centrifuge tubes, various sizes.
- 6.3.2.** Stainless steel spatulas.
- 6.3.3.** Class A volumetric flasks, various sizes.
- 6.3.4.** Glass graduated test tubes.
- 6.3.5.** Disposable pipettes.
- 6.3.6.** Macro pipette controller, various sizes
- 6.3.7.** Pipettes, pipette bulbs.
- 6.3.8.** Aluminum foil squares and plastic weighing dishes for weighing out chemicals.



ANDREW M. CUOMO
Governor

Department of Health

HOWARD A. ZUCKER, M.D., J.D.
Commissioner

SALLY DRESLIN, M.S., R.N.
Executive Deputy Commissioner

7.0. Reagents, Standards and Matrix (Consumables)

- 7.1. Inorganic Chemicals** – Chemicals are obtained from one of the major manufactures such as Sigma-Aldrich, VWR or equivalent. All inorganic chemicals are of reagent grade quality, unless specified in **NYS DOH MML-300** see **Section 7.0**.
- 7.1.1.** Stable solid materials are stored in the laboratory on shelves at room temperature. Concentrated acids are also stored at room temperature in an appropriate cabinet.
- 7.2. Solvents** – All solvents used in sample preparation must be HPLC grade (**NYS DOH MML-300** see **Section 7.0**). Solvents not in use are stored in solvent cabinets.
- 7.2.1.** HPLC grade Acetonitrile (MeCN), Macron or equivalent.
- 7.2.2.** HPLC grade Methanol (MeOH), J.T. Baker or equivalent.
- 7.3. Matrix Reagents**- These materials are common MM excipients or reagents that may be used for matrix evaluation.
- 7.3.1.** Medium-chain triglycerides, Warner Graham (Cat # 812N) or equivalent.
- 7.3.2.** Coconut Oil, Edward International (Cat # S1140-G) or equivalent.
- 7.3.3.** Olive Oil, Sigma-Aldrich (Cat # 01514) or equivalent.
- 7.3.4.** Propylene Glycol, J.T. Baker (Cat # U510-07) or equivalent.
- 7.3.5.** Ethanol (EtOH, Absolute), PHARMCO-AAPER (Cat # 111000200) or equivalent.
- 7.3.6.** New matrices and excipients may also be provided by the ROs for evaluation.
- 7.4. Standards** – The current standards potency analysis are purchased from Cerilliant, Cayman, Restek, Sigma-Aldrich or equivalent (**NYS DOH MML-300** see **Section 7.2**).
- Stock standard solutions or neat materials may be purchased from any vendor. When available, standards/stocks materials are purchased from vendors that can provide NIST traceable standards accompanied by a Certificate of Analysis.
- 7.5. Syringes** – Syringes are obtained from one of the major manufactures such as Hamilton, SGE or equivalent. Manual syringes with fixed or removable needles are stored after cleaning. On arrival in the laboratory, new glassware is cleaned as per **Section 5.2**. Syringes are compliant with relevant standard requirements.
- 7.6. Glassware** – Glassware is obtained from one of the major manufactures of laboratory glassware such as Kimble, Ace Glass, Corning or equivalent. On arrival in the laboratory, new glassware is cleaned as per **Section 5.2**. Glassware is compliant with relevant standard requirements.



ANDREW M. CUOMO
Governor

Department of Health

HOWARD A. ZUCKER, M.D., J.D.
Commissioner

SALLY DRESLIN, M.S., R.N.
Executive Deputy Commissioner

8.0. Preparation of Reagents, Solutions, Standards, Matrices and Samples.

8.1. Standards, SUR and IS are prepared as per NYS DOH MML-300-SOP see **Section 8.0.**

8.2. RO excipient materials and blank matrix (stored as per RO instructions).

8.3. MB, MS, MSD, and sample extract preparation procedure

8.3.1. A direct dilution method is applied for most of the MM products. This method can also be used for extraction of solid material. All the samples are prepared in this manner unless problems are encountered with a specific sample matrix. Any deviations from this sample preparation method are documented and recorded in the data packages. All recoveries are documented and recorded in the data packages.

8.3.2. The amount of MM product to be extracted is based on the RO brand. The weight of matrix used is based on the concentration of cannabinoids in each product to ensure the final concentrations are within the analytical curve. The sample matrix and/or medical marijuana product extract, usually from 10 to 1200 mg, is weighed into a 50-mL centrifuge tube.

8.3.3. The volume of surrogate, 0.005 to 0.040 mL is spiked into the 50-mL centrifuge tube. The amount of surrogate is based on cannabinoid levels in the sample reported by the RO and dilutions needed to ensure the final concentration of the SUR is within the calibration curve.

Typically, a sample that is diluted less than 5-fold will receive 5 μ L of SUR standard stock solution at a concentration of 50 mg/mL as the spike into the sample. Based on the final cannabinoid concentrations, if further dilutions are necessary, the SUR is spiked into the sample at a higher concentration to ensure that the measured concentration is within the calibration range of the SUR standard curve.

8.3.4. For extraction, add 20.0 mL of MeOH to the 50-mL centrifuge tube and, mix well for 30 minutes on a shaker to extract the sample.

8.3.4.1. The final concentration of the cannabinoids in medical marijuana extract must fall within the range of the calibration curve. In some circumstances, an additional dilution of 2 to 20-fold is necessary to analyze the samples. The dilutions are determined based on the concentrations of the cannabinoids in the sample reported by the RO. A larger dilution is needed to bracket high concentration cannabinoids, while a direct injection of the extract or a less diluted sample is required for the analysis of the lower-concentration cannabinoids present in the same sample. Some samples may need to be analyzed twice to measure the primary cannabinoids.

8.3.4.2. Follow NYS DOH MML-300 as per **Section 11.0** for MB, MS and MSD preparation.

8.3.4.3. Sample extracts (**Section 8.3.4**) are stored in a freezer at $\leq -20^{\circ}\text{C}$ until analysis is final. (NYS DOH MML-300 see **Section 9.5**)



8.3.4.4. If necessary, transfer 1 mL of extract into a 2.0 mL centrifuge tube and centrifuge at 12,000 g for 5 min.

8.3.5. Transfer 500 µL IWD preparation @ 10 µg/mL into 2.0 mL HPLC vial (**NYS DOH MML-300 see Section 8.0**).

8.3.6. Transfer 500 µL of diluted sample supernatant prepared (**Section 8.3.4**) into the HPLC vial with IWD (**Section 8.3.5**) and mix well providing a 1:1 ratio.

8.3.7. Follow **NYS DOH MML-300** as per **Section 13.0**, for sample analysis and data reporting.

9.0. Quality Control/Assurance

9.1. Demonstration of Capability (DOC)

9.1.1. All laboratory staff must perform an initial demonstration of capability in using the extraction procedures described in this SOP. The initial DOC must consist of the analysis of four or five extracted spike samples that have been fortified with all analytes of interest at a mid-level concentration. The spiking solution used must be from a source independent from those used to prepare the calibration standards

9.1.2. The initial DOC is performed under the supervision of a trained analyst. The DOC must meet all acceptance criteria, as described in the analytical procedure **NYS DOH MML-300 see section 11.0**, before the analyst may perform the procedure without supervision.

9.1.3. Annually, each analyst who will be performing the extraction method must complete a continuing DOC for each target analyte (**NYS DOH MML-300 Table 1**). The continuing DOC may be completed by one of the following techniques if available:

9.1.3.1. Acceptable performance of a blind sample, such as an external proficiency test.

9.1.3.2. Acceptable performance of an initial DOC as described in **NYS DOH MML-300 see Section 11.0**.

9.1.4. If major changes to the method or instrument are made, or the laboratory/analyst has not performed the method in a twelve (12) month period, each analyst must complete an initial DOC as described in **NYS DOH MML-300 Section 11.0**. Refer to **NYS DOH MML-300 see Section 11.0** for information quality control measures, the applicable acceptance criteria and the corrective actions for nonconforming data. Minor changes to the method are evaluated using the extracted spike, routine samples or the secondary source standard per (**NYS DOH MML-300 see section 11.0**).

9.2. LOD and LOQ

9.2.1. A method specific LOD study must be completed for all target analytes, as listed in **NYS DOH MML -300 Table 1** in a representative matrix, as listed in **Section 7.3**, on each instrument used for analysis of method **NYS DOH MML-300**. The LOD for each cannabinoid is then used to calculate a LOQ for each cannabinoid. Refer to **NYS DOH**



MML-300 see **Section 11.0** and **Section 14.0** for information quality control measures, the applicable acceptance criteria and the corrective actions for nonconforming data.

- 9.2.2. A new LOD study must be verified annually on each instrument for each method, representative matrix and analyte. In addition, LODs are also determined each time there is a significant change in the test method or instrument type.
- 9.2.3. Annually, or when minor changes to the method are made, an LOD study and LOQ verification must be completed as described in **NYS DOH MML 300** see **Section 11.0** to demonstrate continued sensitivity at the LOQ.

9.3. Extraction Batch-Specific Quality Control

- 9.3.1. The batch size consists of a maximum of 20 samples. The following quality control samples must be extracted, where applicable, with each preparation batch of samples at the prescribed frequency:
 - 9.3.1.1. Method Blank, one (1) per extraction batch or every twenty (20) samples.
 - 9.3.1.2. Method Blank Spike, one (1) per batch or every twenty (20) samples.
 - 9.3.1.3. Matrix Spike, one (1) per batch or every twenty (20) samples, if sample is provided.
 - 9.3.1.4. Laboratory Control Sample (LCS) one (1) per batch or every twenty (20) samples.
- 9.3.2. Refer to the analytical procedure (**NYS DOH MML-300**) for information on the quality control measures, the applicable acceptance criteria and the corrective actions for nonconforming data.

9.4. Analytical Batch-Specific Quality Control

- 9.4.1. Refer to analytical procedure (**NYS DOH MML-300**) for information on quality control measures, applicable acceptance criteria and corrective actions for nonconforming data.

10.0. Data Acquisition, Reduction, Analysis and Calculations

- 10.1. Not applicable; refer to the appropriate analytical procedure (**NYS DOH MML-300**).

11.0. Sample Collection, Preservation, Handling and Storage

- 11.1. Medical Marijuana Products from Registered Organizations are received, handled, verified and documented ensuring method regulatory and Accreditation Body requirements are met.
- 11.2. Follow instructions provided by the RO for storage prior to sample extraction.
- 11.3. Prior to analysis, the extracts are stored in a freezer at $\leq -20^{\circ}\text{C}$ unless otherwise noted (**NYS DOH MML-300** see **Section 9.0.**)
- 11.4. Cannabinoids are light sensitive so samples and must therefore be protected from the light.



ANDREW M. CUOMO
Governor

Department of Health

HOWARD A. ZUCKER, M.D., J.D.
Commissioner

SALLY DRESLIN, M.S., R.N.
Executive Deputy Commissioner

12.0. Waste Management/Pollution Prevention

- 12.1. Minimize solvent, chemical, reagent, and standard use whenever possible to reduce the amount of hazardous waste generated.
- 12.2. Dispose of solvent waste in an appropriate solvent waste container, properly labeled.
 - 12.2.1. All other solvents are separated into two categories, chlorinated and non-chlorinated, and are disposed of in red, 5-Gallon solvent cans.
- 12.3. Dispose of non-hazardous water waste in the laboratory sink followed by flushing with tap water.
- 12.4. Dispose of glassware in appropriately labeled boxes. Be sure that, whenever possible, the glass has been thoroughly rinsed and is contaminant-free before disposal.
- 12.5. Consult federal, state and local regulations for additional information or for information on the disposal of products not described in this method.

13.0. References

- 13.1. Measurement of Phytocannabinoids in Medical Marijuana by HPLC-PDA. **NYS DOH MML-300.**
- 13.2. Definition and Procedure for the Determination of the Method Detection Limit--Revision 1.11 Environmental Protection Agency, 40 CFR (7-1-95 Edition) Part 136, Appendix B.
- 13.3. *Norgestrel*; MSDS No. N2260 [Online]; Sigma-Aldrich: Saint Louis MO, September 03, 2014 <http://www.sigmaaldrich.com/catalog/AdvancedSearchPage.do>